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# 11 COCAINE, COCAETHYLENE AND BENZOYLECGONINE QUANTITATION AND CONFIRMATION BY SPE AND GCMS

## 11.1 Summary

11.1.1 Biological samples are extracted using United Chemical Technologies® solid phase extraction columns. The extracts are concentrated, treated with a derivatizing reagent and injected into the GCMS for confirmation and quantitation by selected ion monitoring.

## 11.2 Specimen Requirements

11.2.1 2 mL of whole blood, biological fluids or tissue homogenates.

#### 11.3 Reagents And Standards

- 11.3.1 Cocaine, cocathylene and benzoylecgonine, 1 mg/mL
- 11.3.2 Cocaine-d<sub>3</sub> and/or benzoylecgonine-d<sub>3</sub>, 100 μg/mL
- 11.3.3 Concentrated Acetic Acid
- 11.3.4 Methanol
- 11.3.5 Hexane
- 11.3.6 Dichloromethane
- 11.3.7 Isopropanol
- 11.3.8 Acetonitrile
- 11.3.9 Concentrated Ammonium Hydroxide
- 11.3.10 N-Methyl-N-([tert-butyldimethyl-silyl)trifluoroacetamide (MTBSTFA) or N,O bis (Trimethylsilyl) trifluoroacetamide (BSTFA) with 1% TMCS
- 11.3.11 Ethyl acetate

#### 11.4 Solutions, Internal Standards, Calibrators, Controls

- 11.4.1 1 M Acetic Acid. Add 100-200 mL  $dH_2O$  to a 1L volumetric flask. Add 57.5mL glacial acetic acid. QS to volume with  $dH_2O$ .
- 11.4.2 Dichloromethane/isopropanol, 80:20 (v:v): mix 800 mL dichloromethane and 200 mL isopropanol.
- 11.4.3 When using UCT CleanScreen® SPE Extraction columns, either sodium or potassium phosphate buffer may be used. However, the same buffer (sodium or phosphate) must be used throughout the duration of the procedure.
- 11.4.4 0.1M Potassium Phosphate Buffer, pH 6.0. Weigh out 13.61 g of KH<sub>2</sub>PO<sub>4</sub> and transfer into a 1 L volumetric flask containing approximately 800 mL of dH<sub>2</sub>O. Adjust the pH of the above solution to 6.0 by the addition of 5 M potassium hydroxide while stirring. QS to volume with dH<sub>2</sub>O. Solution can also be purchased (e.g. Fisher).

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11.4.5	0.1M Sodium Phosphate Buffer, Ph 6.0. Weigh out 1.70g $Na_2HPO_4$ and 12.14g $NaH_2PO_4 \cdot H_2O$ and transfer to L volumetric flask containing approximately 800 mL $dH_2O$ . Adjust the pH of the above solution to 6.0 by the addition of 5 M sodium hydroxide. QS to volume with $dH_2O$ . Solution can also be purchased (e.g. Fisher).		
11.4.6	Dichloromethane/Isopropanol/Ammonium Hydroxide elution solvent: Mix 78 mL dichloromethane with 20 mL isopropanol. Mix well. In hood, add 2 mL of concentrated NH <sub>4</sub> OH. Mix gently.		
11.4.7	Working standard solutions		
	11.4.7.1	$100\mu\text{g/mL}$ cocaine, cocaethylene and benzoylecgonine: Pipet solution of cocaine, cocaethylene and benzoylecgonine into a 10 with methanol.	
	11.4.7.2	$10\mu\text{g/mL}$ cocaine, cocaethylene and benzoylecgonine: Pipet 1. cocaine, cocaethylene and benzoylecgonine into a 10 mL volumethanol.	
	11.4.7.3	Working internal standard solution, $10~\mu g/mL$ cocaine- $d_3$ and/or the $100~\mu g/mL$ stock solutions of cocaine- $d_3$ and/or benzoylecge QS to volume with methanol.	
11.4.8	The following are examples of acceptable procedures for the preparation of calibrators. Other quantitative diluti may be acceptable to achieve similar results.		
	11.4.8.1	Cal 1: $2.5 \text{ mg/L}$ : $500 \mu\text{L}$ of $10 \mu\text{g/mL}$ working standard + $2.0 \mu\text{m}$	blank blood
	11.4.8.2	Cal 2: $1.0 \text{ mg/L}$ : $200 \mu\text{L}$ of $10 \mu\text{g/mL}$ working standard + $2.0 \mu\text{L}$	blank blood
	11.4.8.3	Cal 3: 0.5 mg/L: $100 \mu\text{L}$ of $10 \mu\text{g/mL}$ working standard + $2.0 \mu$	blank blood
	11.4.8.4	Cal 4: $0.25$ mg/L: $50 \mu L$ of $10 \mu g/mL$ working standard + $2.0$	blank blood
	11.4.8.5	Cal 5: $0.05$ mg/L: $10 \mu L$ of $10 \mu g/mL$ working standard + $2.0$	blank blood
	11.4.8.6	Cal 6: $0.02 \text{ mg/L}$ : $4 \mu \text{L}$ of $10 \mu \text{g/mL}$ working standard + $2.0 \text{ b}$	lank blood
	11.4.8.7 Cal 7: 0.01 mg/L: $2 \mu L$ of $10 \mu g/mL$ working standard + 2.0 blank blood		
11.4.9	Prepare the first standard by adding 200 $\mu$ L of the 100 $\mu$ g/mL to 4.8 mL blank blood to give a final concent of 4 mg/L.		ank blood to give a final concentrati
	11.4.9.1	Cal 1: 4.0 mg/L: 2 mL of 4 mg/L standard	
	11.4.9.2	Cal 2: 1.5 mg/L: 750µL of 4 mg/L standard + 1.25mL blank bl	lood
	11.4.9.3 Cal 3: 0.5 mg/L: 250μL of 4 mg/L standard + 1.75mL blank blood		
	11.4.9.4	Cal 4: 0.25 mg/L: 125µL of 4 mg/L standard + 1.85mL blank	blood
	11.4.9.5	Cal 5: 0.10 mg/L: 50µL of 4 mg/L standard + 1.9mL of blank	blood

11.4.9.6 Cal 6: 0.05 mg/L:  $25\mu L$  of 4 mg/L standard + 2 mL of blank blood

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	11.4.10	11.4.9.7 Cal 7: 0.02 mg/L: 10μL c Controls	of 4 mg/L standard + 2 mL of blank bl	ood		
		11.4.10.1 Negative blood control. B cocaethylene or benzoyled	Blood bank blood (or comparable) dete gonine.	rmined not to contain cocaine,		
		11.4.10.2 QAS Toxicology Control: 0.1 mg/L cocaine and cocaethylene and 1.0 mg/L benzoylecgonine				
		11.4.10.3 In house control is prepare cocaethylene and benzoyle	ed from a different lot number or a diffection ecgonine.	erent manufacturer of cocaine,		
11.5	Appara	Apparatus				
	11.5.1	Agilent GC/MSD, Chemstation software, compatible computer & printer				
	11.5.2	Test tubes, 16 x 125 mm round bottom, screw cap tubes, borosilicate glass with Teflon caps				
	11.5.3	Test tubes, 16 x 125 mm round bottom tubes, borosilicate glass				
	11.5.4	Test tubes, 16 x 114 mm (10 mL) glass tubes, conical bottom				
	11.5.5	Centrifuge capable of 2,000 – 3,000 rpm				
	11.5.6	Cleanscreen® Extraction Cartridges (ZSDAU020) from United Chemical Technologies (200 mg columns)				
	11.5.7	Solid phase extraction manifold				
	11.5.8	Vortex mixer				
	11.5.9	Heating block				
	11.5.10	Evaporator/concentrator				
	11.5.11	GC autosampler vials and inserts				
	11.5.12	HP GC/MSD				
		11.5.12.1 Acquisition Mode:	SIM			
		11.5.12.2 Cocaine:	<u>303</u> , 198, 272	Alternate ions: <u>182</u> , 303, 198		
		11.5.12.3 Cocaine-d <sub>3</sub> :	<u>306,</u> 185			
		11.5.12.4 Cocaethylene:	<u>317</u> , 196, 272	Alternate ions: <u>82</u> , 196, 317		
		11.5.12.5 Benzoylecgonine:	MTBSTFA ions <u>403</u> , 282, 346	BSTFA ions: <u>240</u> , 361, 256		
		11.5.12.6 Benzoylecgonine-d <sub>3</sub> :	MTBSTFA ions <u>406</u> , 285	BSTFA ions: <u>85</u>		
		11.5.12.7 Column:	HP 5MS 25 m x 0.25 mm x 0.25 μm			
		11.5.12.8 Detector Temperature:	280° C			

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11.5.12.9 Instrument conditions may be changed to permit improved performance. 11.5.12.9.1 Oven Program

Equilibration time: 0.50 minutes
Initial temp: 130° C
Initial time: 1 minutes
Ramp: 17° C/min
Final Temp: 280° C
Final Time: 7 minutes
Run Time: 17 minutes

11.5.12.9.2 Inlet

Mode: Splitless
 Temperature: 250° C
 Injection volume: 1.0 μL

• Purge Time: ON at 2.0 minute

#### 11.6 Procedure

- 11.6.1 Label clean 16 x 125 mm screw cap tubes accordingly, negative, calibrators, control(s) and case sample IDs.
- 11.6.2 Pipet 2 mL of blank blood, calibrators, controls and case sample bloods, fluids or tissue homogenates in appropriately labeled tubes.
- 11.6.3 Add 50 µL internal standard into all tubes and vortex.
- 11.6.4 Add 4 mL deionized water to each tube. Vortex briefly and let stand for 5 minutes.
- 11.6.5 Centrifuge at approx 2000 rpm for 10 minutes.
- 11.6.6 Add 2 mL of pH 6 phosphate buffer to remaining supernatant.
- 11.6.7 Condition the solid phase extraction columns. Throughout the SPE procedure, it is important not to permit the SPE sorbent bed to dry, unless specified. If necessary, add additional solvent/buffer to re-wet.
  - 11.6.7.1 Add 3 mL hexane to each column and aspirate on vacuum manifold
  - 11.6.7.2 Add 3 mL methanol to each column and aspirate on vacuum manifold.
  - 11.6.7.3 Add 3 mL dH<sub>2</sub>O and aspirate.
  - 11.6.7.4 Add 1 mL of 0.1 M pH 6.0 phosphate buffer and aspirate
- 11.6.8 Without delay, pour specimens into appropriate SPE columns. Elute from cartridges with ~ 1-2 mL/ minute flow.
- 11.6.9 Wash the solid phase extraction columns:
  - 11.6.9.1 Add 3 mL dH<sub>2</sub>O and aspirate at  $\leq$  3 inches of mercury.
  - 11.6.9.2 Repeat the dH<sub>2</sub>O wash a second time.

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- 11.6.9.3 Wash with 2.0 mL 1.0 M acetic acid and aspirate.
- 11.6.9.4 Add 3 mL methanol and aspirate.
- 11.6.9.5 Add 2 mL hexane and aspirate. Dry the columns at > 10 inches of Hg for at least 10 minutes.
- 11.6.10 Wipe the SPE column tips with Kimwipes®. Place labeled 10 mL conical test tubes in the manifold test tube rack. Be sure SPE column tips are in the designated conical tube.
- 11.6.11 Elute drugs by adding 3 mL of freshly prepared dichloromethane/isopropanol/ammonium hydroxide solution to each column. Collect eluate by gravity drain (no vacuum).
- 11.6.12 Evaporate to dryness at approximately 40° C under nitrogen.
- 11.6.13 Derivatize specimens:
  - 11.6.13.1 Derivatize by adding 50µL ethyl acetate and 50 µL BSTFA and heat for 15 minutes at 55° C OR
  - 11.6.13.2 Derivatize by adding 50  $\mu$ L of MTBSTFA and heat for 30 minutes at 85° C. Then add 50 $\mu$ L of ethyl acetate.
- 11.6.14 Transfer to GC microvials. Inject 1.0 µL on GC/MS in the SIM mode.

### 11.7 Calculations

- 11.7.1 Calculate the concentrations by interpolation of a linear plot of the response curve based on peak height (or area) ratios (using the target ions listed under GCMS conditions) versus calibrator concentration.
- 11.7.2 Qualifier ion ratio range. The qualifier ion ratio range is calculated by determining the mean  $\pm$  20% (or 2 SD) ion ratio from all calibrators used in the calibrations curve. Each drug has two qualifier ions ratios and each internal standard has one.

## 11.8 Quality Control

11.8.1 See Toxicology Quality Guidelines

#### 11.9 References

- 11.9.1 United Chemical Technologies, Inc. Clean Screen® solid phase extraction procedure for cocaine and benzoylecgonine from whole blood.
- 11.9.2 Spiehler, Vina R. and Reed, Dwight (1985) Journal of Forensic Science, 30:4, 1003-1011.
- 11.9.3 Crouch, Dennis J., et al. (1995) Journal of Analytical Toxicology, 19, 352-358.